ON THE ANTIVIRAL ACTIVITY OF DIFFUSOMYCIN (OXAZOLOMYCIN)

E. TONEW, M. TONEW, U. GRÄFE, P. ZÖPEL

Central Institute for Microbiology and Experimental Therapy, 0-6900 Jena, F. R. Germany

Received August 10, 1990; revised July 2, 1991

Summary. - The effect of the β -lactone antibiotic diffusomycin (oxazolomycin) was investigated against vaccinia (Lister), herpes simplex type 1 (Kupka), influenza A (WSN; H₁N₁), and Coxsackie A9 viruses. Diffusomycin reduced significantly the plaque formation of enveloped DNA and RNA viruses by more than 90 % in the range of the maximally tolerated dose. As could be shown with vaccinia virus, the antiviral action was not caused by virucidal effect on virions or by interaction with virus adsorption and penetration. In one-step growth cycle assays diffusomycin prevented the replication of herpes simplex type 1, vaccinia and influenza A viruses in a dose-dependent manner. The replication of influenza A viruses was blocked immediately after addition of the compound during zero to six hr p.i. Partial reversibility of the antiviral action was established by washing off the antibiotic from chicken embryo cells (CEC) infected with influenza A virus. Finally, replication of Coxsackie A9 virus was not inhibited by diffusomycin. Electronoptical studies revealed a reduced synthesis of HSV-1 nucleocapsids in dependence on the concentration of the compound.

Key words: diffusomycin; vaccinia virus; herpes simplex virus type 1; influenzavirus A (WSN); in vitro inhibition; virus replication

Introduction

Numerous antibiotics have been investigated on their antiviral activities in vitro. However, only few of them combine properties such as high antiviral activity and low toxicity in vivo. Diffusomycin was isolated from cultures of Streptomyces albus (JA 3453) and found identical with oxazolomycin (Mori et al., 1985) when comparing the relevant high-field ¹H and ¹³C NMR data (Gräfe et al., 1988a, 1988b). The drug possesses promising antitumour and antibacterial properties (Mori et al., 1985), but no information has been given with regard to any other biological activities so far, neither was its antiviral action investigated. In this paper we report on the antiviral activity of diffusomycin against influenza A, vaccinia and herpes simplex viruses in vitro.

Materials and Methods

Substance. Diffusomycin (oxazolomycin) was isolated and purified to homogenity as described (Gräfe et al., 1988a); its chemical structure is shown in Fig. 1.

Cell cultures. Chicken embryo cells (CEC) were cultivated in Hanks solution with 10 % Parker 199 medium and 5 % calf serum (Tonew and Tonew, 1969). RH cells (continuous human kidney cell line) were grown in a medium consisting of one part Eagle's MEM and one part Hanks solution with 0.5 % lactalbuminhydrolysate with 5 % calf serum (Tonew and Glück, 1986). Human fibroblast cells were cultured with Eagle's MEM containing 5 % neonatal calf serum (SIFIN, Berlin).

Viruses. Vaccinia virus (strain Lister), herpes simplex virus type 1 (HSV-1) (strain Kupka), influenza virus A/WSN (H_1N_1) were propagated in CEC and RH cells, Coxsackievirus type A9 in human fibroblast cells.

Maximally tolerated dose (MTD). MTD was determined by microscopic observations as the compound concentration which caused no morphological alterations of cultured cells up to day 5 after treatment.

Antiviral activity. Assay for virucidity, influence on adsorption and penetration processes have been described earlier (Tonew and Tonew, 1971).

Plaque reduction test. The test was carried out without and with the test compound in different concentrations in an agar overlay as described by Tonew and Tonew (1969).

One step growth cycle experiments (OSGE). Using a m.o.i. of 20 PFU/cell and after washing off the nonadsorbed virus (three times), the infectious virus yield was determined according to Reed and Muench (1938) at different drug concentrations and in its absence by the end of the replication cycle (vaccinia virus and HSV-1 by 24 hrs, influenza virus WSN by 8 hrs p.i.).

Reversal of the antiviral activity. After infection with a m.o.i. of about 20 PFU/cell of influenza virus A/WSN in the presence of 15.75 μ g/ml diffusomycin further procedures were carried out according to Tonew et al. (1975).

Microtitration assay. Coxsackie virus type A9 in different dilutions was checked against the inhibitor with MTD as well as with decreased concentrations in human fibroblast cells grown on titertek plates (Flow). The titre of untreated as well as antibiotic treated virus-infected cell cultures was determined according to Reed and Muench (1938).

Electron-optical examinations. Investigations were performed during OSGE in CEC infected with HSV type 1 in the presence of the antibiotic. Cell pellets were embedded and cut using the Reichert Wien ultramicrotome apparatus as reported (Zöpel et al., 1973) and after contrasting viewed in Siemens Elmiscope 1 at 80 kV.

Results

The MTD of diffusomycin for CEC and RH cells was 125 μ g/ml on day 5, which caused 88 to 100 % decrease of the plaque number of the three enve-

Fig. 1
Structure formula of diffusomycin (oxazolomycin)

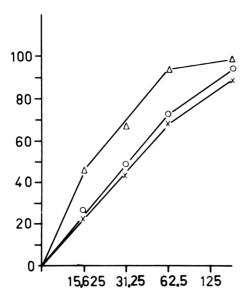


Fig. 2
Plaque reduction of HSV-1 (△), influenza
(○) and vaccinia (x) viruses in CEC in dependence on the concentrations of diffusomycin (72 hr p.i.)

Ordinate: plaque reduction in per cent; abscissa: concentration in μ g/ml

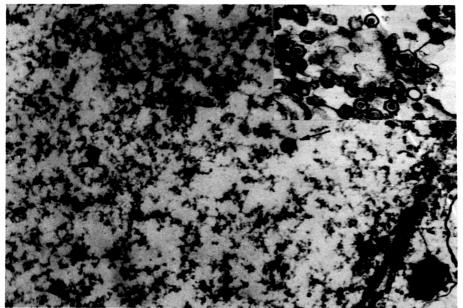


Fig. 3 HSV-1-infected CEC treated with 31.25 μ g/ml diffusomycin. Occurence of a few nucleocapsids in the nucleus

Insert: nucleus of untreated virus-infected control cell. Magnification 60,000 x

Table 1. Inhibition of virus replication by diffusomycin in one-step growth cycle experiments using RH cells and CEC (m.o.i. 20 TCID₅₀/cell)

1			
c virus type	CEC	2.37 (>99.9) 2.7 (>99.9) 3.93 (>99.9) (>99.9)	8.37
Herpes simplex virus type	RH cells	2.14 (>99.9) 2.66 (>99.9) 3.94 (>99.9) 5.66 (99.7)	8.14
Vaccinia virus Titre in log ₁₀ TCID ₅₀ /ml	CEC	2.7 (>99.9) 2.92 (>99.9) 4.92 (99) 5.96 (89)	6.92
Vaccini Titre in log ₁₀	RH cells	3.53 (>99.9) 4.2 (>99.7) 5.27 (96.3) 5.87 (85.2)	6.7
Influenza A virus	CEC	3.7 (>99.9) 4.2 4.2 (>99.9) 5.63 (>99.9) (99.9)	8.87
Influenz	RH cells	4.2 (>99.9)*) 4.2 (>99.9) 5.04 (>99.9) 6.2 (99.7)	8.7
Concentration	diffusomycin in µg/ml	125 62.5 31.25 15.62	0

*) In parenthesis: per cent inhibition

Table 2. Lack of activity of t	penetration of vaccinia v	_	e adsorption and

Concentration	Cell-free virus Titre in log ₁₀ TCID ₅₀ /ml after incubation at 37 °C		PFU after treatment during		
in μg/ml			adsorption	penetration	
	4 hrs	20 hrs		with immui	without noserum
125 0	6.7 6.87	5.92 6.18	121 137	114 126	129 141

loped viruses checked in CEC. A significant reduction was yet observed with 31.25 μ g/ml diffusomycin (Fig. 2).

The drug in concentrations of 125 and 62.5 μ g/ml reduced the infectious virus yield of influenza A, vaccinia and herpes simplex type 1 viruses during OSGE in both human and chicken cells by more than 99 % (Table 1).

The antibiotic showed neither virucidal effect nor an influence on virus adsorption and penetration processes as could be demonstrated with vaccinia viruses (Table 2). The mode of action was further studied only with influenza virus WSN. Infectious virus production was inhibited after application of the compound at zero as well as at 4 and 6 hr p.i. (Table 3). Removal of the antibiotic after 3 hr partially restored virus replication in a time depending manner. Haemagglutinin synthesis started with a few hours delay when the antibiotic treatment was shorter than 4 hr (Table 4). The antibiotic did not interfere with replication of Coxsackie virus type A9 in human fibroblast cells as tested by

Table 3. Antiviral activity of diffusomycin (125 μ g/ml) against influenza virus A in CEC when added at 0.4 and 6 hr p.i.

Substance	HAU/ml		Virus yield in log ₁₀ TCID ₅₀ /ml		
application at hrs	with diffus	without somycin	with diffus	without omycin	
0	4	4	2.92	3.37	
4	4	8	3.39	3.86	
6	64	128	6.87	7.37*	

^{*} Titre of the untreated virus control at 8 hr p.i. was 8.87 log₁₀ TCID₅₀/ml. Titres were determined by 8 hr after addition of the compound.

Table 4. Reversion of the antiviral activity of diffusomycin (15.62 μ g/ml) in dependence on the time of removal of the antibiotic

Duration		Hou	urs after virus infe	ction	
of compound treatment in hrs	0	4	8 HAU/ml	24	48
2	n.t.	2	16	128	256
3	n.t.	4	8	64	64
4	n.t.	4	8	16	32
O (Control)	< 2	4	128	256	512

n.t. not tested

microtitration assays. The electron-optical investigations during replication of herpes simplex virus type 1 support the effect of the compound on virus replication. No virus particles could be observed in the presence of $62.5 \,\mu\text{g/ml}$ diffusomycin, whereas after application of $31.25 \,\mu\text{g/ml}$ only a few nucleocapsids appeared (Fig. 3).

Discussion

The β -lactone antibiotic diffusomycin (oxazolomycin) may represent a new structural type of antiviral agent. The antibiotic suppressed the replication of vaccinia, herpes simplex type 1, and influenza A viruses during OSGE both in CEC and RH cells. The mode of antiviral action of membranotropic antibiotics, in general, could involve inhibition of virus uncoating as an early stage of virus replication (Korant *et al.*, 1984). But diffusomycin apparently inhibited later stages of influenza A replication (about 4 to 6 hr p.i.) suggesting other yet unidentified ability of the drug acting during maturation and/or virus release. The absence of susceptibility of Coxsackie virus A9 appears to be an interesting fact which will promote further studies towards elucidation of the mode of antiviral action of diffusomycin.

Acknowledgement. The skillful technical assistance of Mrs. B. Gumpert is gratefully appreciated.

References

Gräfe, U., Fleck, W. F., Schade, W., Wiesner, J., Tonew, E., and Tresselt, D. (1988a): Verfahren zur Herstellung eines Antibiotikums. Patentschrift WP C12 P, 315 1915. Gräfe, U., Dornberger, K., and Fleck, W. F. (1988b): Approaches to new microbial metabolites with nonclassical mode of action. *Progr. Ind. Microbiol.* 27, 113-138.

- Korant, B. D., Lonberg-Holm, K., and LaColla, P. (1984): Picorna viruses and Togaviruses: Targets for Design of Antivirals, pp. 61-97. In E. De Clerqc and R. T. Walker (Eds): Target for the Design of Antiviral Agents, Plenum Publishing Corporation.
- Mori, T., Takahashim, K., Kashiwabara, M., and Lemura, D. (1988): Structure of Oxazolomycin, a novel β -lactone antibiotic. *Tetrahedron Lett.* 26, 1073-1076.
- Reed, J., and Muench, H. (1938): A simple method of estimating fifty per cent endpoints. *Amer. J. Hyg.* 27, 493-497.
- Tonew, E., and Tonew, M. (1969): Die Wirkung von Cycloheximid auf Abortus-Virus der Schafe, Sindbis-Virus und andere Viren in Screening-Untersuchungen. Zbl. Bakt. Hyg. I. Orig. 211, 437-444.
- Tonew, E., Tonew, M., Eckardt, K., Thrum, H., and Gumpert, B. (1975): Streptovirudins-new antibiotics with antiviral activity. The antiviral spectrum and the inhibition of Newcastle disease virus in cell cultures. *Acta virol.* 19, 311-317.
- Tonew, M., and Glück, B. (1986): Interferonsensitivität verschiedener Zellkulturen. J. basic Microbiol. 26, 173-189.
- Tonew, M., and Tonew, E. (1971): Antivirale Wirkung vom Imidazol-derivaten. I. Die Hemmung der Vermehrung des Mengovirus in FL-Zellen. Arch. ges. Virusforsch. 33, 319-329.
- Zöpel, P., Eckardt, K., and Tonew, E. (1973): Elektronen-opticshe Untersuchungen zur Wirkung des Streptomycetenantibiotikums Borrelidin auf das Pseudorabiesvirus. Z. Allg. Mikrobiol. 13, 711-722.